

DRIMANE DERIVATIVES FROM *DRIMYS BRASILIENSIS*

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Key Word Index—*Drimys brasiliensis*; Winteraceae; drimane sesquiterpenes; confertifolin; 1 β -*p*-coumaroyloxyvaldiviolide; 1 β -*p*-coumaroyloxyvaldiviolide.

Abstract—*Drimys brasiliensis* gave confertifolin and two new drimane derivatives identified as 1 β -*p*-coumaroyloxyvaldiviolide and 1 β -*p*-coumaroyloxyvaldiviolide.

INTRODUCTION

Drimys sensu stricto is a small South American genus of four species, two of which have been investigated and provided the first examples of what are known as drimane sesquiterpenes [1–4]. We now report the isolation from *Drimys brasiliensis* Miers of confertifolin (1) and the new drimane derivatives 1 β -*p*-coumaroyloxyvaldiviolide (2) and 1 β -*p*-coumaroyloxyvaldiviolide (3a).

RESULTS AND DISCUSSION

Confertifolin (1) has been isolated previously from *D. confertifolia* Phil. and in much better yield from *D. winteri* Forst. It also accompanies polygodial (4) in the pungent *Polygonum hydropiper* L. (Polygonaceae) [5–7]. Mp, IR and NMR data of our material compared well with those in the literature [2–4, 8].

Compound 2, C₂₈H₂₄O₅ (HRMS), was a *p*-coumarate derivative as evidenced by the typical signals of H-2', H-3', H-5' and H-6' in its ¹H NMR spectrum (Table 1), the signals of C-1'–C-7' in its ¹³C NMR spectrum (Table 2) and the loss of C₉H₈O₃ on EIMS. Partial structure A could also be deduced from the ¹H and ¹³C NMR spectra. One of the two aldehyde functions (C-12) was α,β -unsaturated (H-12, *s*, δ 9.32, C-12, *d*, δ 193.21), the second (C-11) was normal (H-11, *d*, δ 9.82, C-11, *d*, δ 200.18) and adjacent to a methinyl group (H-9, *s* (*br*), δ 3.34, C-9, *d*, δ 60.35) whose proton was further allylically coupled to a vinylic proton (H-7 at δ 7.09, C-7 at δ 153.37) β to the α,β -unsaturated aldehyde. In turn H-7 and also a very shielded proton (H-5) appeared as a doublet of doublets at δ 1.44 and were coupled to the protons (H-6) of a methylene group appearing as a complex multiplet centred at δ 2.48.

Partial structure A and the presence of three methyl singlets in the ¹H NMR spectrum indicated that 2 was a polygodial derivative. That the ester function was equatorial and located at C-1 or C-3 of the drimane skeleton was deduced from the magnitude (11 and 4 Hz) and number of the coupling constants involving the doublet of doublets at δ 4.89. The NOE difference spectrum (Table 3) showed an appreciable interaction between H-1 and H-9 and located the coumarate at C-1. It also showed that H-1, H-5 and H-9 were all axial, thus

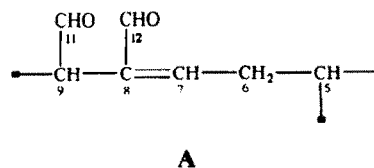
establishing the complete stereochemistry of 2. That H-5 was axial and α was also shown by the coupling constants (11 and 6 Hz).

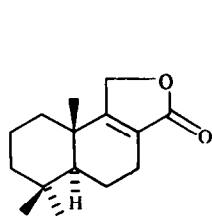
The ¹H NMR data for 2 compared well with those of polygodial (4) where applicable. The 250 MHz ¹H NMR spectrum of the latter has been reported recently [9]. The H-11 signal of 2 was considerably further downfield than in the case of polygodial and was more in keeping with that of 9-epipolygodial [10], but the NOEs and the value of *J*_{9,11} ruled this out. Obviously H-11 of 2 was deshielded by the ester function on C-1.

Comparison of the ¹H and ¹³C NMR spectra (Tables 1 and 2) of the second new compound, C₂₈H₂₄O₆ (HRMS), with those of 2 permitted its formulation as the 1 β -*p*-coumaroyloxy derivative 3a of valdiviolide (5). The latter is a minor constituent of *Drimys winteri* [4]. Oxidation of the α,β -unsaturated aldehyde of 2 to a carboxyl and lactal formation between C-11 and C-12 accounted for the extra oxygen atom of the empirical formula, the slight upfield shift of H-7 and the significant upfield shifts of C-11 (to δ 116.77), C-12 (to δ 170.17) and H-11 (to δ 5.78). H-11 experiences a predictable paramagnetic shift to δ 6.56 on conversion to the diacetate 3b. Since the coupling constants exhibited by 3a, b compare well in most respects with those of 2, the stereochemistry of 3a is the same as that of 2.

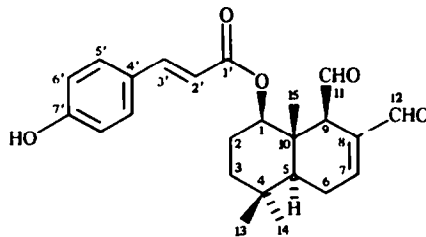
EXPERIMENTAL

Extraction of *Drimys brasiliensis*. Aerial parts of *D. brasiliensis* Miers. (7.5 kg) were collected by Dr. Hermogenes de Freitas Leitão Filho in Campos do Jordão, São Paulo State, Brazil, in

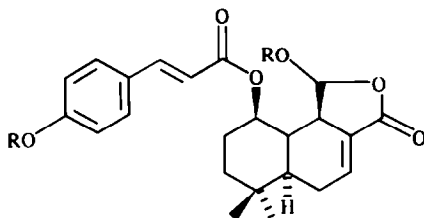




1

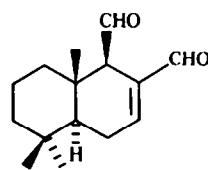


2

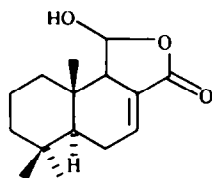


3a R = H

3b R = Ac



4



5

March 1974. The pulverized plant was extracted with hexane-EtOAc (5:1) to give 37 g crude extract which was chromatographed over 276 g silica gel, 500 ml fractions being eluted as follows: 1–13 (hexane-EtOAc, 49:1), 14–28 (hexane-EtOAc, 24:1), 29–51 (hexane-EtOAc, 15.6:1), 52–81 (hexane-EtOAc, 11.5:1), 82–91 (hexane-EtOAc, 9:1), 92–104 (hexane-EtOAc, 7.3:1), 105–108 (hexane-EtOAc, 5.2:1), 109–114 (hexane-EtOAc, 4:1), 115–120 (hexane-EtOAc, 3:1), 121–126 (hexane-EtOAc, 2.5:1), 127–131 (hexane-EtOAc, 1.5:1), 132–138 (EtOAc) and 139–147 (EtOH). The crystals from fraction 39 were recrystallized from hexane-EtOAc (49:1) to give 30 mg **1**, 147–150°; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 2902, 1760, 1740, 1670, 1430, 1000; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1755 (br), 1670 cm^{-1} ; $^1\text{H NMR}$: δ 4.74 (*ddd*, $J = 17.3, 3$ Hz) and 4.66 (*ddd*, $J = 17, 3.5, 1.8$ Hz, AB system of H-11 allylically coupled to H-7), 2.40 *dd* (br), $J = 17, 6$ Hz) and 2.16 (*m*, AB system of H-7), 1.91 (*dd* (br), $J = 13, 7$ Hz, probably H-6a), 1.17, 0.95 and 0.92 (3 Me).

The solid material (12 g) from fr. 118–133 was rechromatographed on silica gel (84 g); the material eluted with hexane-EtOAc (2.3:1) gave 32 mg **3a**, mp 164–166°;

IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3450, 3320, 1710, 1690, 1630, 1605, 1590, 1510; MS m/z (rel. int.): 412 [$\text{M}]^+$ (1.4), 394 (2.2), 384 (3.9), 265 (2.1), 246 (3.2), 230 (9.0), 220 (26.0), 205 (13.9), 191 (11.0), 164 (18.8), 150 (5.0), 147 (100) [Calc. for $\text{C}_{28}\text{H}_{24}\text{O}_6$: M_r , 412.1884. Found: M_r (MS) 412.1874]. Other significant peaks in the HRMS were at m/z (composition, rel. int.): 394 ($\text{C}_{24}\text{H}_{26}\text{O}_5$, 0.4), 384 ($\text{C}_{23}\text{H}_{28}\text{O}_5$, 0.6), 366 ($\text{C}_{23}\text{H}_{26}\text{O}_4$, 0.2), 248 ($\text{C}_{15}\text{H}_{20}\text{O}_3$, 0.5), 220 ($\text{C}_{14}\text{H}_{20}\text{O}_2$, 11.9), 164 ($\text{C}_9\text{H}_8\text{O}_3$, 20.5), 150 ($\text{C}_9\text{H}_8\text{O}_2$, 9.2), 147 ($\text{C}_9\text{H}_7\text{O}_2$, 100). Acetylation in the usual fashion ($\text{Ac}_2\text{O}-\text{C}_5\text{H}_5\text{N}$) gave **3b**, MS m/z (rel. int.): 496 [$\text{M}]^+$ (2.9), 454 (14.2), 437 (2.7), 394 (6.0), 230 (74), 220 (11.8), 215 (10.6), 189 (17.8), 164 (14.4), 147 (100).

Fractions 126–127 gave 54 mg **2**, mp 201–204°; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3450 (br), 1725 (br), 1680 (br); MS m/z (rel. int.): 396 [$\text{M}]^+$ (0.6), 368 (1.3), 232 (0.8), 204 (23.5), 175 (4.2), 164 (10.6), 147 (100). [Calc. for $\text{C}_{24}\text{H}_{18}\text{O}_5$: M_r , 396.1937. Found: M_r (MS), 396.1941].

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Table 1. ^1H NMR spectra of compounds 2 and 3a, b (270 MHz, CDCl_3)

H	2	3a*	3b
1 α	4.89 <i>dd</i> (11, 4)	4.74 <i>dd</i>	4.70 <i>dd</i>
2 α	1.71 <i>dq</i> (4, 12)	1.68 <i>dq</i>	1.68 <i>dq</i>
2 β	1.82 <i>dq</i> (12, 4)	1.88 <i>dq</i>	1.94 <i>dq</i>
3 α , β †	1.58 <i>m</i>	1.58 <i>m</i>	1.58 <i>m</i>
5 α	1.44 <i>dd</i> (11, 6)		1.5 <i>m</i>
6 α		2.45 <i>ddt</i> (20, 5.5, 3.5)	2.52 <i>ddt</i>
6 β	2.48 <i>m</i>	2.22 <i>ddd</i> (20, 12, 4, 3)	2.25 <i>m</i>
7	7.09 <i>br</i> ($W_{1/2}$ = 9 Hz)	6.86 (part obsc)	6.96 <i>q</i> (3.5)
9 α	3.34 <i>br</i> ($W_{1/2}$ = 7 Hz)	2.59 <i>m</i>	2.86 <i>ddt</i> (5.5, 4, 3.5)
11	9.82 <i>d</i> (2.8)	5.78 <i>d</i> (5.5)	6.56 <i>d</i> (5.5)
12	9.35	—	—
13‡	1.09	1.00	1.04
14‡	1.03	0.97	0.94
15‡	0.98	0.94	0.96
2'	6.23 <i>d</i> (16)	6.28 <i>d</i>	6.26 <i>d</i>
3'	7.59 <i>d</i> (16)	7.42 <i>d</i>	7.65 <i>d</i>
5'†	6.87 <i>d</i> (8)	6.89 <i>d</i>	7.14 <i>d</i>
6'†	7.39 <i>d</i> (8)	7.42 <i>d</i>	7.55 <i>d</i>
Ac‡	§		2.32 ¶, 1.88**

* Plus 2 drops DMSO.

† Intensity two protons.

‡ Intensity three protons.

§ In absence of D_2O , OH singlet at δ 5.96.|| In absence of D_2O , OH signals at δ 9.16 and δ 2.45, H-11 signal split into triplet.

¶ On C-7'.

** On C-11.

Table 2. ^{13}C NMR spectra of compounds 2 and 3a (67.89 MHz)

C	2 (deutero-acetone)	3a (CD_3OD)
1	81.59 <i>d</i>	81.77 <i>d</i>
2	23.36 <i>t</i> *	25.83 <i>t</i> *
3	39.91 <i>t</i>	40.55 <i>t</i>
4	33.40 <i>s</i>	33.69 <i>s</i>
5	49.21 <i>d</i>	obsc. by solvent
6	24.99 <i>t</i> *	25.18 <i>t</i> *
7	153.37 <i>d</i>	137.26 <i>d</i>
8	140.92 <i>s</i>	129.71 <i>s</i>
9	60.55 <i>d</i>	59.41 <i>s</i>
10	42.33 <i>s</i>	39.70 <i>s</i>
11	200.18 <i>d</i>	116.77 <i>d</i> †
12	193.21 <i>d</i>	170.17 <i>s</i>
13	32.75 <i>q</i>	32.71 <i>q</i>
14	22.26 <i>q</i>	21.54 <i>q</i>
15	10.95 <i>q</i>	10.11 <i>q</i>
1'	166.54 <i>s</i>	168.73 <i>s</i>
2'	115.94 <i>d</i>	116.21 <i>d</i>
3'	145.62 <i>d</i>	145.96 <i>d</i>
4'	127.06 <i>s</i>	127.39 <i>s</i>
5'	131.04 <i>d</i>	131.19 <i>d</i>
6'	116.69 <i>d</i>	116.77 <i>d</i> †
7'	160.62 <i>s</i>	161.15 <i>s</i>

* Assignments interchangeable.

† Signals superimposed.

Table 3. ^1H NOE difference spectrum of 2a

Saturation	Observed NOE (%)
H-1 α	H-5 α (4.1)
	H-9 α (10.6)
H-5 α	H-1 α (10.3)
	H-9 α (8.6)
H-9 α	H-1 α (11.3)
	H-5 α (7.3)

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